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24.09.2004



INVESTOR IN PEOPLE

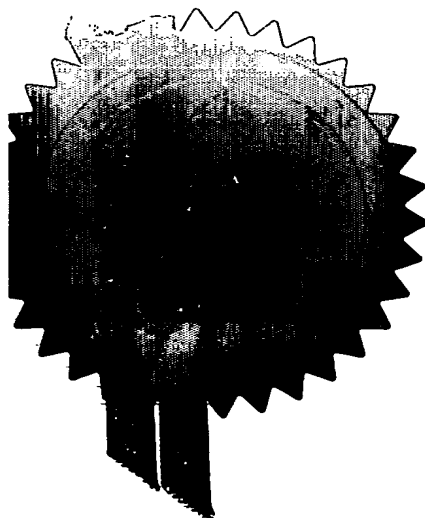
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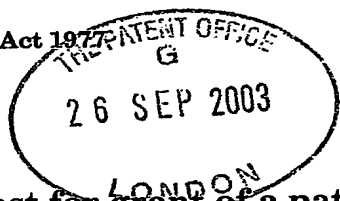
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# 1/77

## Request for grant of a patent

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The Patent Office

Cardiff Road  
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1.	Your reference	4-33364P1		
2.	Patent application number (The Patent Office will fill in this part)	0322612.3		
3.	Full name, address and postcode of the or of each applicant (underline all surnames)	NOVARTIS AG LICHTSTRASSE 35 4056 BASEL SWITZERLAND Patent ADP number (if you know it) 7125487005 If the applicant is a corporate body, give the country/state of its incorporation SWITZERLAND		
4.	Title of invention	Organic compounds		
5.	Name of your agent (If you have one) "Address for service" in the United Kingdom to which all correspondence should be sent (including the postcode)	Craig McLean Novartis Pharmaceuticals UK Limited Patents and Trademarks Wimbleshurst Road Horsham, West Sussex RH12 5AB Patents ADP number (if you know it) 07181522002		
6.	If you are declaring priority from one or more earlier patent applications, give the country and the date of filing of the or of each of these earlier applications and (if you know it) the or each application number	Country	Priority application number (if you know it)	Date of filing (day/month/year)
7.	If this application is divided or otherwise derived from an earlier UK application, give the number and the filing date of the earlier application	Number of earlier application		Date of filing (day/month/year)
8.	Is a statement of inventorship and of right to grant of a patent required in support of this request? (Answer 'Yes' if: a) any applicant named in part 3 is not an inventor, or b) there is an inventor who is not named as an applicant, or c) any named applicant is a corporate body. (see note (d))	Yes		

## Patents Form 1/77

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Continuation sheets of this form

Description 7

Claim(s) 3

Abstract

Drawing(s)

- 
10. If you are also filing any of the following, state how many against each item.

Priority documents

Translations of priority documents

Statement of inventorship and right to grant of a patent (*Patents Form 7/77*)

Request for preliminary examination and search (*Patents Form 9/77*) 1

Request for substantive examination (*Patents Form 10/77*)

Any other documents  
(please specify)

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11.

I/We request the grant of a patent on the basis of this application

Signature

Date



Craig McLean

25<sup>th</sup> September 2003

- 
12. Name and daytime telephone number of person to contact in the United Kingdom

Mr. Trevor Drew

01403 323069

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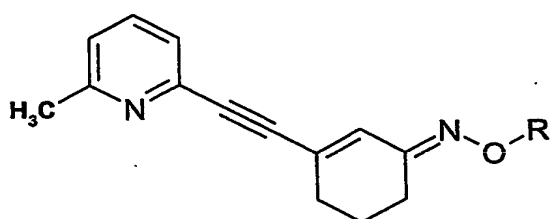
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Organic Compounds

The present invention relates to novel pyridylacetylene derivatives, their preparation, their use as radiotracers/markers and compositions containing them.

More particularly the invention provides a compound of formula I



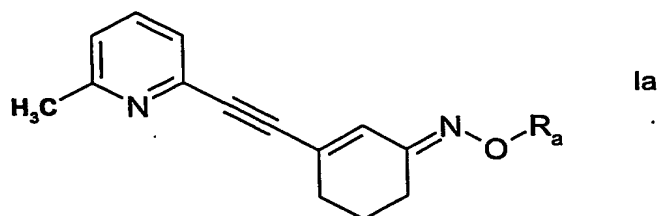
wherein

R is  $^{11}\text{CH}_3$ ,  $(^3\text{H})_3\text{C}$ ,  $(\text{CH}_2)_n^{123}\text{I}$ ,  $(\text{CH}_2)_n^{76}\text{Br}$  or  $(\text{CH}_2)_n^{18}\text{F}$ , n being 1, 2, 3 or 4

in free base or acid addition salt form.

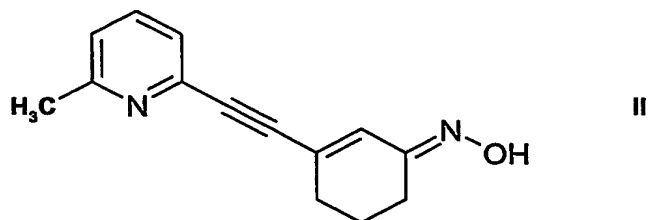
In a further aspect, the invention provides a process for the production of the compounds of formula I and their salts, comprising the steps of

a) for the production of a compound of formula Ia



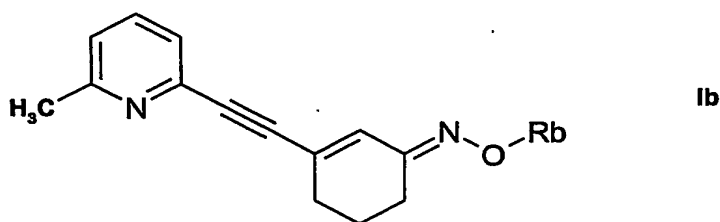
wherein  $\text{R}_a$  is respectively  $^{11}\text{CH}_3$  or  $(^3\text{H})_3\text{C}$ , reacting the compound of formula II

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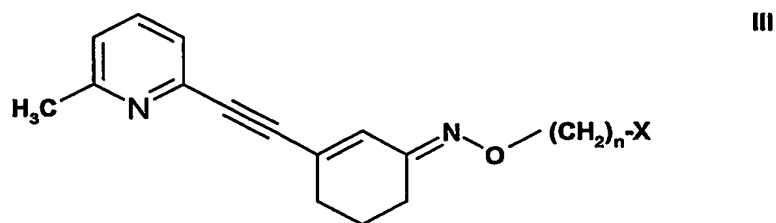


with respectively  $^{11}\text{CH}_3\text{I}$  or  $\text{C}(^3\text{H})_3\text{I}$ , in the presence of a base, or

b) for the production of a compound of formula Ib



wherein Rb is respectively  $(\text{CH}_2)_n^{18}\text{F}$ ,  $(\text{CH}_2)_n^{123}\text{I}$  or  $(\text{CH}_2)_n^{76}\text{Br}$ , reacting a compound of formula III



wherein n is as defined above and X is OTs or OMs, with respectively  $^{18}\text{F}^\ominus$ ,  $^{123}\text{I}^\ominus$  or  $^{76}\text{Br}^\ominus$ , or reacting the compound of formula II with a compound of formula IV



wherein X and Rb are as defined above,

and recovering the resulting compound of formula I in free base form or in form of an acid addition salt.

The reactions can be effected according to known methods, for example as described in the Examples.

Working up the reaction mixtures and purification of the compounds thus obtained may be carried out in accordance to known procedures.

Acid addition salts may be produced from the free bases in known manner, and vice-versa.

The starting materials of formulae II, III and IV are known or may be obtained in analogous manner to known procedures, e.g. as described in the Examples.

Compounds of formula I in free base or acid addition salt form, hereinafter referred to as agents of the invention, exhibit valuable properties as histopathological labeling agents, imaging agents and/or biomarkers, hereinafter "markers", for the selective labeling of the metabotropic glutamate receptor subtype 5 (mGlu5 receptor).

More particularly the agents of the invention are useful as markers for labeling the central and peripheral mGlu5 receptors *in vitro* or *in vivo* (see Example 5-7).

The agents of the invention are therefore useful, for instance, for determining the levels of receptor occupancy of a drug acting at the mGlu5 receptor, or diagnostic purposes for diseases resulting from an imbalance or dysfunction of mGlu5 receptors, and for monitoring the effectiveness of pharmacotherapies of such diseases.

In accordance with the above, the present invention provides an agent of the invention for use as a marker for neuroimaging.

In a further aspect, the present invention provides a composition for labeling brain and peripheral nervous system structures involving mGlu5 receptors *in vivo* and *in vitro* comprising an agent of the invention.

In still a further aspect, the present invention provides a method for labeling brain and peripheral nervous system structures involving mGlu5 receptors *in vitro* or *in vivo*, which comprises contacting brain tissue with an agent of the invention.

The method of the invention may comprise a further step aimed at determining whether the agent of the invention labeled the target structure. Said further step may be effected by observing the target structure using positron emission tomography (PET) or single photon emission computed tomography (SPECT), or any device allowing detection of radioactive radiations.

The following examples illustrate the invention.

**Example 1: 3-(6-Methyl-pyridin-2-ylethynyl)-cyclohex-2-enone O-[<sup>11</sup>C-methyl]-oxime**

3-(6-Methyl-pyridin-2-ylethynyl)-cyclohex-2-enone O-[<sup>11</sup>C-methyl]-oxime is synthesized by reacting [<sup>11</sup>C]-MeI with the sodium salt of 3-(6-methyl-pyridin-2-ylethynyl)-cyclohex-2-enone oxime (1mg) in dry DMF (400 µl). [<sup>11</sup>C]-MeI is introduced via a slow stream of helium and when addition is completed the reaction mixture is heated to 120°C for 10 min. Product purification is accomplished by reversed phase HPLC using a C-18 µ-Bondapak column (7.8x300mm) and a mobile phase consisting of CH<sub>3</sub>CN/0.1% H<sub>3</sub>PO<sub>4</sub> (70/30) at a flow rate of 5 ml/min. The retention time of the desired product is between 6 and 7 min.

3-(6-Methyl-pyridin-2-ylethynyl)-cyclohex-2-enone O-[<sup>11</sup>C-methyl]-oxime is formulated in a solution containing polysorbatum (2%), ethanol (10%) and saline (0.9%).

LogD = 2.5 (determined using the classical shake-flask method).

The starting materials are prepared as described hereafter:

**a) 3-(6-Methyl-pyridin-2-ylethynyl)-cyclohex-2-enone**

A solution of 2-ethynyl-6-methyl-pyridine (702 mg, 6 mmol), 3-bromo-cyclohex-2-enone (1.26 g, 7.2 mmol), bis-(triphenylphosphine)-palladium-dichloride (168 mg, 0.24 mmol), copper(I) iodide (93 mg, 0.48 mmol), triethylamine (4.8 ml, 34.4 mmol) in 12ml DMF is heated to 55°C for 1 h. After that time the solution is diluted with ethyl acetate (500 ml) and washed with sat aq. NaHCO<sub>3</sub> (1 X 150 ml). The water phase is extracted with ethyl acetate (1 X 150 ml) and the combined organic phases are dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuo. The residue (1.88 g) is purified on column chromatography (silica gel, eluent hexane/ethyl acetate 3:1 v/v) and the fractions containing the desired compound are collected and

concentrated in vacuo to yield 1.05 g (yield = 82 %) of the title compound as a light yellow oil.

b) 3-(6-Methyl-pyridin-2-ylethynyl)-cyclohex-2-enone oxime

A solution of 3-(6-methyl-pyridin-2-ylethynyl)-cyclohex-2-enone (422 mg, 2 mmol) and hydroxylamine hydrochloride (278 mg, 4 mmol) in pyridine (20 ml) is stirred for 17 h at RT. After that time the solvent is evaporated in vacuo. The residue is dissolved in ethyl acetate (300 ml) and washed with sat NaHCO<sub>3</sub> (1 X 50 ml). The water phase is extracted with ethyl acetate (1 X 50 ml). The combined organic phases are dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuo. The residue (0.45 g) is purified on column chromatography (Silica gel, eluent hexane/ethyl acetate 2:1 v/v) and the fractions containing the desired compound are collected and concentrated in vacuo to yield 0.192 g (yield = 42 %) of the title compound as light yellow crystals, m.p. 166-168°C.

**Example 2: 3-(6-Methyl-pyridin-2-ylethynyl)-cyclohex-2-enone O-[tri(<sup>3</sup>H)-methyl]-oxime**

The title compound can be prepared by reacting 3-(6-methyl-pyridin-2-ylethynyl)-cyclohex-2-enone oxime with [<sup>3</sup>H]-MeI (0.5 equivalent) in the presence of K<sub>2</sub>CO<sub>3</sub> in DMF at 100°C for 180 min, followed by a purification by reversed phase chromatography.

**Example 3: 3-(6-Methyl-pyridin-2-ylethynyl)-cyclohex-2-enone O-(2-[<sup>18</sup>F-fluoro]-ethyl)-oxime**

The sodium salt of the 3-(6-methyl-pyridin-2-ylethynyl)-cyclohex-2-enone oxime (2mg) is reacted in dry DMF (400 µl) with [<sup>18</sup>F]-2-fluoro-ethyltosylate (obtained from ethyleneditosylate and [<sup>18</sup>F]-KF-Kryptofix complex) at 100°C for 10min. The reaction mixture is purified through a tC-18 Sep-Pak cartridge, and the fractions containing the desired product are further purified by a semi-preparative reversed phase HPLC using a C18 Bondclone column (300x7.8 mm) and a mobile phase consisting of CH<sub>3</sub>CN/0.01M H<sub>3</sub>PO<sub>4</sub> (70/30) at a flow rate of 4 ml/min. The fraction containing the product (retention time between 12 and 13 min) is passed through a tC-18 Sep-Pak cartridge and eluted with 1 ml of ethanol. This ethanolic solution is buffered with 0.15M phosphate buffer to give after sterile filtration an isotonic and injectable solution.



**Example 4: 3-(6-Methyl-pyridin-2-ylethynyl)-cyclohex-2-enone O-[<sup>18</sup>F-fluoro]-methyl)-oxime**

The sodium salt of the 3-(6-methyl-pyridin-2-ylethynyl)-cyclohex-2-enone oxime (2mg) is reacted in dry DMF (400 µl) with [<sup>18</sup>F]CH<sub>2</sub>OTf at 100°C for 30min. After an initial purification by passing the reaction mixture through a tC-18 Sep-Pak cartridge, 3-(6-methyl-pyridin-2-ylethynyl)-cyclohex-2-enone O-[<sup>18</sup>F-fluoro]-methyl-oxime was finally purified by a semi-preparative reversed phase HPLC using a C18 Bondclone column (300x7.8 mm) and a mobile phase consisting of CH<sub>3</sub>CN/0.01M H<sub>3</sub>PO<sub>4</sub> (70/30) at a flow rate of 4 ml/min. The product is formulated in analogy to 3-(6-methyl-pyridin-2-ylethynyl)-cyclohex-2-enone O-(2-[<sup>18</sup>F-fluoro]-ethyl)-oxime (example 3).

**Example 5: K<sub>i</sub>/IC<sub>50</sub> determination (binding assay)**

*In vitro*, the affinity for the mGlu5 receptor is determined using a radioligand displacement technique as described by Gasparini et al, Bio. Med. Chem. Let. 2002, 12, 407-409. 3-(6-methyl-pyridin-2-ylethynyl)-cyclohex-2-enone O-methyl-oxime shows an IC<sub>50</sub> of 8 nM (Hill coefficient 1.08; 95 % confidence intervals: 6.0-10.0 nM) for the displacement of [<sup>3</sup>H]-2-methyl-6-((3-methoxyphenyl)ethynyl)-pyridine from membrane of L-tk cells stably expressing the human mGlu5 receptor (Daggett et al, Neuropharm. 1995, 34:871-886). Using the Cheng-Prusoff equation, a K<sub>i</sub> of 4 nM is calculated (radioligand concentration used for the assay: 2 nM).

**Example 6: Organ and brain structure distribution**

Two groups of male adult Sprague-Dawley rats weighing 250-300 g are used for the biodistribution studies. The first group (n = 3) serves as the control group and the second group (n = 3) serves as the blockade group. Each animal received 250-300 pmol (0.6-40 MBq) of 3-(6-methyl-pyridin-2-ylethynyl)-cyclohex-2-enone O-[<sup>11</sup>C-methyl]-oxime via a lateral tail vein. The blockade group is co-injected with 2-methyl-6-((3-methoxyphenyl)ethynyl)-pyridine (1 mg/kg) whereas the control group (n=3) receives a corresponding volume of 0.9% NaCl. The animals are sacrificed 30 min post-injection. Organ or brain regions such as hippocampus, striatum, cortex and cerebellum are removed and measured in a gamma-counter. The tissue distribution is expressed as percentage of injected dose per gram wet tissue (% ID/g organ).

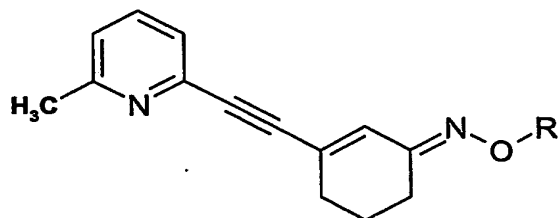
**Example 7: Results of the brain distribution study with 3-(6-methyl-pyridin-2-ylethynyl)-cyclohex-2-enone O-[<sup>11</sup>C-methyl]-oxime**

The table below displays the percentage of the injected dose normalized per gram of tissue. K1, K2, K3 are individual values for control animals. B1, B2, B3 are individual values for animals co-injected with 2-methyl-6-((3-methoxyphenyl)ethynyl)-pyridine.

Organ	K1	K2	K3	B1	B2	B3
Hippocampus	0.16154	0.25440	0.21264	0.05184	0.06195	0.05972
Striatum	0.22564	0.30685	0.28164	0.06495	0.07158	0.06487
Cortex	0.14849	0.18316	0.16909	0.04964	0.05582	0.05431
Cerebellum	0.03322	0.03925	0.03699	0.04052	0.03608	0.03421
Midbrain	0.07703	0.10559	0.09091	0.03876	0.04700	0.04422
Restbrain	0.05519	0.06520	0.06070	0.03832	0.04762	0.04383
Whole brain	0.11123	0.13659	0.12999	0.04580	0.05118	0.04951

**CLAIMS**

1. A compound of formula I



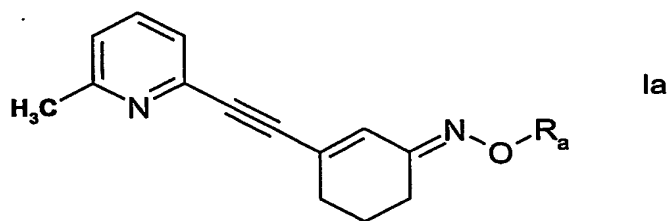
wherein

R is  $^{11}\text{CH}_3$ ,  $(^3\text{H})_3\text{C}$ ,  $(\text{CH}_2)_n^{123}\text{I}$ ,  $(\text{CH}_2)_n^{76}\text{Br}$  or  $(\text{CH}_2)_n^{18}\text{F}$ , n being 1, 2, 3 or 4

in free base or acid addition salt form.

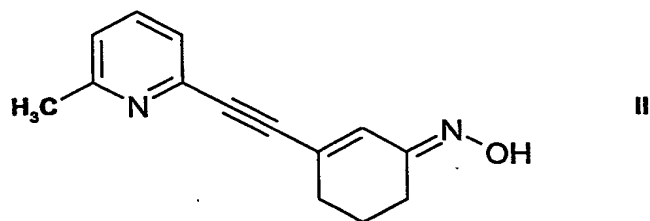
2. A process for the production of a compounds of formula I as defined in claim 1, or a salt thereof, comprising the step of

a) for the production of a compound of formula Ia



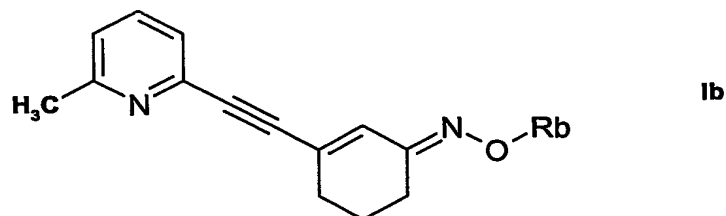
wherein  $\text{R}_a$  is respectively  $^{11}\text{CH}_3$  or  $(^3\text{H})_3\text{C}$ , reacting the compound of formula II

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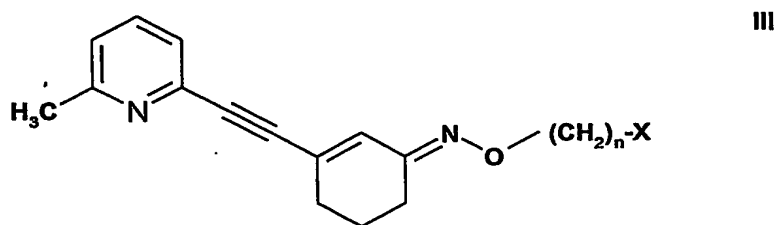


with respectively  $^{11}\text{CH}_3\text{I}$  or  $\text{C}(^3\text{H})_3\text{I}$ , in the presence of a base, or

b) for the production of a compound of formula Ib



wherein Rb is respectively  $(\text{CH}_2)_n^{18}\text{F}$ ,  $(\text{CH}_2)_n^{123}\text{I}$  or  $(\text{CH}_2)_n^{76}\text{Br}$ , reacting a compound of formula III



wherein n is as defined in claim 1 and X is OTs or OMs, with respectively  $^{18}\text{F}^\ominus$ ,  $^{123}\text{I}^\ominus$  or  $^{76}\text{Br}^\ominus$ , or reacting the compound of formula II with a compound of formula IV



wherein X and Rb are as defined above,

and recovering the resulting compound of formula I in free base form or in form of an acid addition salt.

3. A compound of formula I as defined in claim 1, in free base or acid addition salt form, for use as a marker for neuroimaging.
4. A composition for labeling brain and peripheral nervous system structures involving mGlu5 receptors *in vivo* and *in vitro* comprising a compound of formula I as defined in claim 1, in free base or acid addition salt form.
5. A method for labeling brain and peripheral nervous system structures involving mGlu5 receptors *in vitro* or *in vivo*, which comprises contacting brain tissue with a compound of formula I as defined in claim 1, in free base or acid salt form.

**PCT/EP2004/010743**

